



Available online at <https://int-scientific-journals.com>

International Scientific Journals



IJEISR (2019) Vol.3–no.2

<https://int-scientific-journals.com/ijeisr>

# Achieving Reliable Outcomes by Adopting Proper Quality Samples

*Andrea M. Steinriede*

*Department of Legal Medicine, Christian-Albrechts-University of Kiel, Arnold-Heller-Str. Kiel, Germany*

*Email: a.steinriede@uk-sh.de*

## **Abstract**

The study discusses using certain methods that were tested with sets of standards and has given reliable results. Always there is a potential for laboratory results to become wrong results. Therefore, performing quality control using quality samples on the medical devices existed in the laboratory will be with great benefits as well as ensuring the perfect performance of the equipment. In this paper, some important highlights will be reported. Some definitions of the associated terms will be discussed. However, some well-known papers will be referred to in order to provide some valuable information about this topic. In conclusion, performing a quality sample provide correct information and enables the medical staff to make preventive or corrective measures early.

Doi: [10.31219/osf.io/xryge](https://doi.org/10.31219/osf.io/xryge)

© 2019 The Authors. Published by ISJ.

This is an open access article under the CC BY 4.0 license (<https://creativecommons.org/licenses/by-nc-nd/4.0/>)

Keywords: *Reliable Result, Quality Parameters, Reliable Sample.*

---

## Introduction

Recently, the interest in the medical sector has increased and technology has invaded all its fields. However, the doctor, the nurse and the patient are also keen to obtain laboratory results that are of high accuracy and do not expose the patient to the suffering of repeated sample work again. Not only this, but also reducing the chances of waste of resources in the labs, preservation of property and reduce its expenses. From here, the term “control sample” is emerged recently [2,4,8].

A control sample is well-known as controls and is defined as any type of any well-known forensic sample and its objective is to ensure analyses, as well as tests, are properly performed yielding to reliable results [7]. In addition, control samples are of different types in terms of composition, identification, source and type [9,11]. Researchers and scientists illustrate the importance of these samples in two approaches:

- i. Assure that a test run is valid.
- ii. Assure that results are reliable [3].

In the laboratory technology, control samples should be utilized in the execution of each inspection. Moreover, control samples should be addressed in the precise same technique as the test samples and are used to validate the test run [2,6]. Recently, each laboratory should employ its standards for run acceptance established on instructions from the manufacturer's kit (i.e. the negative and positive control provided with the kit) insert and in-lab validation of an external quality control sample [8].

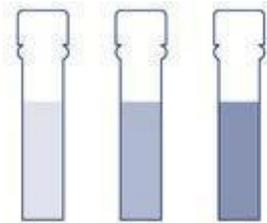
## Analyses Workflow

Before the delivering of the first lab test, the journey starts. It starts with making a calibration to the devices. Some devices might be programmed in advance to make a self-test every morning at a specific time and some devices ask the technician to make the calibration because it may require some reagents. Each workflow involving different consecutive steps that modify depending on the sample type and iterate the sample until it is contingent to extract its information and turn it into actionable results. Challenges are within every step promptly regarded to the sample type and analysis technologies, and at each step, there is a high risk for things to go in the wrong direction. Therefore, performing accurate quality control is of high importance to ensure quality results and reliable interpretation [10,12,14].

In fact, there are four significant quality parameters which enable to characterize the quality of what is called the nucleic acid samples and these four parameters determined depending on the downstream application [1]. The four parameters are: quantity, purity, size and sequence. Before considering the features embedded in these parameters, it is important to note the importance of the stability in these parameters [3,8,11,13]. Indeed, various variation in these parameters may manipulate the experiments and impact the data quality as well as results interpretation. The variation in these parameters even small one could drive the assay to become a failure [5].

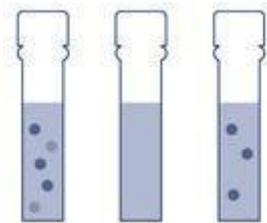
The influence of the four significant parameters on downstream applications should be with great effort. This influence implies evaluating these parameters, performing quick and simple quality control of various measures and increase the chances of success on the assay [8,11].

In general, quantity implies enough material to perform the assay. It is the first step to ensure the correct results. The technician should be able to determine the accurate sample quantification. However, the accidental estimation errors might impact the downstream assays. Figure 1 below shows three bottles with different materials to perform the assay and it is easily seen that first bottle on the right is the one with the suitable quantity.



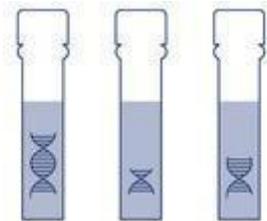
*Fig 1: Quantity Parameter*

Another parameter impacts the results of an assay is the purity. It is the parameter that ensures that the sample does not contain any contaminants. As a consequence, impurities have a high risk to interpose with the downstream application. Figure 2 below illustrates this parameter and it is obvious that the bottle in the middle is the one with no contaminants.



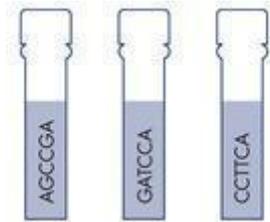
*Fig 2: Purity Parameter*

An equally significant aspect is the size parameter. The size parameter can be defined as the right size of the distribution of the sample and ensuring that the sample is not degraded. Accordingly, estimating the size of the nucleic acid size distribution affords insights into the integrity of the sample, in addition, this action will distinguish any poor quality or any degraded samples that are not well performed for analysis. Figure 3 below shows the size parameter



*Fig 3: Size Parameter*

At last, the sequence is the fourth parameter. Performing this parameter enables the technician to assure that the sample is the with the right genotype. It is the final step; it is considered as the primary validation step. This parameter ensures that the technician is performing the full credit to obtain right results interpretation. Figure 4 below illustrates this parameter.



*Fig 4: Sequence Parameter*

A lot of risks face the technicians of a laboratory when neglecting the principles of quality control in general and quality samples specifically. Performing these actions in the morning demand not more than a few minutes and does not cost a lot of money. On the contrary, performing these actions has the possibility to ensure that the technicians' task is performing the samples of the highest quality so, in other words, it might save the money as well as the time.

Validating everything associated with a quality control in its right way in the lab might ensure in advance stages a profitable investment. Also, it is very important to use high-performance tools to ensure the validity of the four significant parameters.

Nowadays, researchers and scientists are much more sensitive to detection technologies than before. But we should also consider that these technologies are not necessarily more firm in terms of tolerating variations into the quality of a sample.

## **Conclusions**

Monitoring the success of any experiment stands in front of a lot of parameters. The control sample characterizes the ultimate success step and it is important to ensure the existence of the four parameters before performing the assay.

## References

1. C. KOEBERL, Neutron Activation Analysis, in: A. MARFUNIN (Ed.), Higher Mineralogy, Springer Verlag, 1991. [Google Scholar]
2. G. A. WANDLESS, Radiochemical neutron activation analysis of geologic materials, U.S. Geol. Surv. Bull., 1770 (1987). [Google Scholar]
3. J. W. JACOBS, R. L. KOROTEV, D. P. BLANCHARD, L. A. HASKIN, J. Radioanal. Chem., 40 (1977) 93. [Google Scholar]
4. C. KOEBERL, F. KLUGER, W. KIESL, J. Radioanal. Nucl. Chem., 112 (1987) 481. [Google Scholar]
5. C. KOEBERL, E. H. HAGEN, Geochim. Cosmochim. Acta, 53 (1989) 937. [Google Scholar]
6. C. KOEBERL, J. Trace Microprobe Techn., 6 (1988) 501. [Google Scholar]
7. C. KOEBERL, W. KIESL, Meteoritics, 21 (1986) 420. [Google Scholar]
8. K. GOVINDARAJU, Geostand. Newsl., 11 (1984) 203. [Google Scholar]
9. E. JAROSEWICH, R. S. CLARKE, Jr., J. N. BARROWS (Ed.), The Allende Meteorite Reference Sample, Smithsonian Contrib. Earth Sci., Vol. 27, 1987, p. 49. [Google Scholar]
10. C. KOEBERL, Proc. NIPR Symp. Antarct. Meteorites, 1 (1988) 122. [Google Scholar]
11. C. KOEBERL, P. H. WARREN, M. M. LINDSTROM, B. SETTEL, T. FUKUOKA, Proc. NIPR Symp. Antarct. Meteorites, 2 (1989) 15. [Google Scholar]
12. C. KOEBERL, G. KURAT, F. BRANDSTÄTTER, Proc. NIPR Symp. Antarct. Meteorites, 3 (1990) 3. [Google Scholar]
13. C. KOEBERL, G. KURAT, F. BRANDSTÄTTER, Geochim. Cosmochim. Acta, 55 (1991) 3073. [Google Scholar]
14. C. KOEBERL, G. KURAT, F. BRANDSTÄTTER, Proc. NIPR Symp. Antarct. Meteorites, 4 (1991) 33. [Google Scholar]



This work is licensed under a Creative Commons Attribution 4.0 International License.